

# In-situ, real-time measurement of glucose during mammalian cell culture

Sarvani Manne, Shriram Kaliannan Chandramohan, Jean-François P. Hamel  
(Contact: Dr. Hamel, jhamel@mit.edu)

Presented at :  
BPI Conference  
Boston, MA  
October 5-7 2016

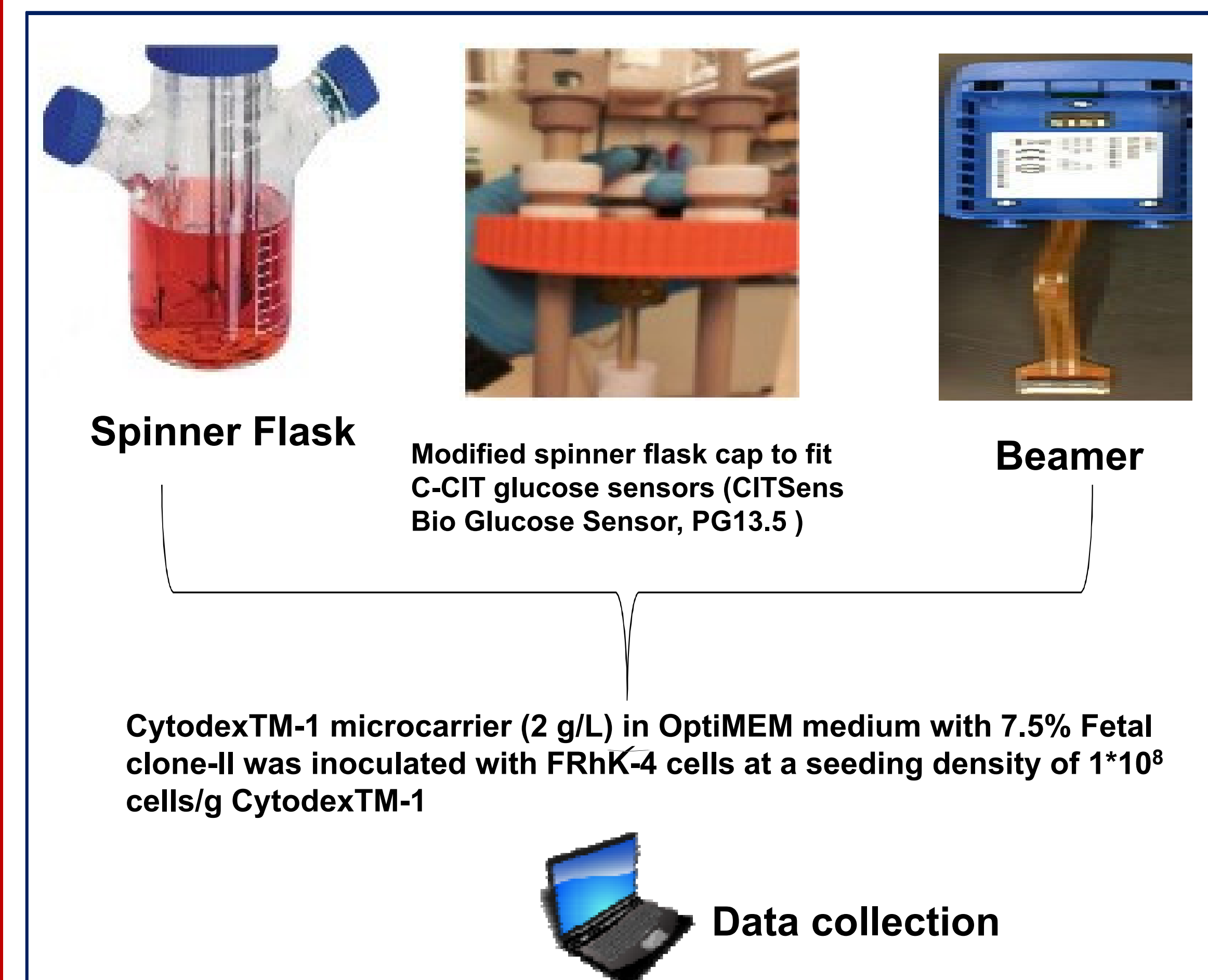
Department of Chemical Engineering, Massachusetts Institute of Technology, Cambridge, MA 02139

## ABSTRACT

A novel online glucose sensor (C-CIT Sensors AG) was employed for monitoring glucose levels in microcarrier cell culture, and compared to offline analysis with a biochemical analyzer (BioProfile® Flex). The C-CIT glucose sensor is based on immobilized enzyme technology and is connected to a wireless transmitter. The format of the sensor enables its inclusion into the T-flask, spinner flask and bioreactor. In this study, FRhK-4 cells were grown over Cytodex-1 micro carriers in OptiMEM medium with 7.5% v/v Fetal Clone-2 serum. The cells were grown in 500-mL spinner flask for 23 days. Glucose trends generated from the online sensor were found to be consistent with punctual offline analysis.

## MATERIALS AND METHODS

CITSens Bio Glucose Sensor, PG13.5 was integrated in glass spinner flask (500 mL Corning Proculture®).



### Instrument Set-up:

- Sensor was conditioned for 8 hours before calibration
- *In-situ* sensor was calibrated for 1 h before inoculation

**Agitation:** 45 RPM (increased to 60 RPM as the micro carriers started settling down)

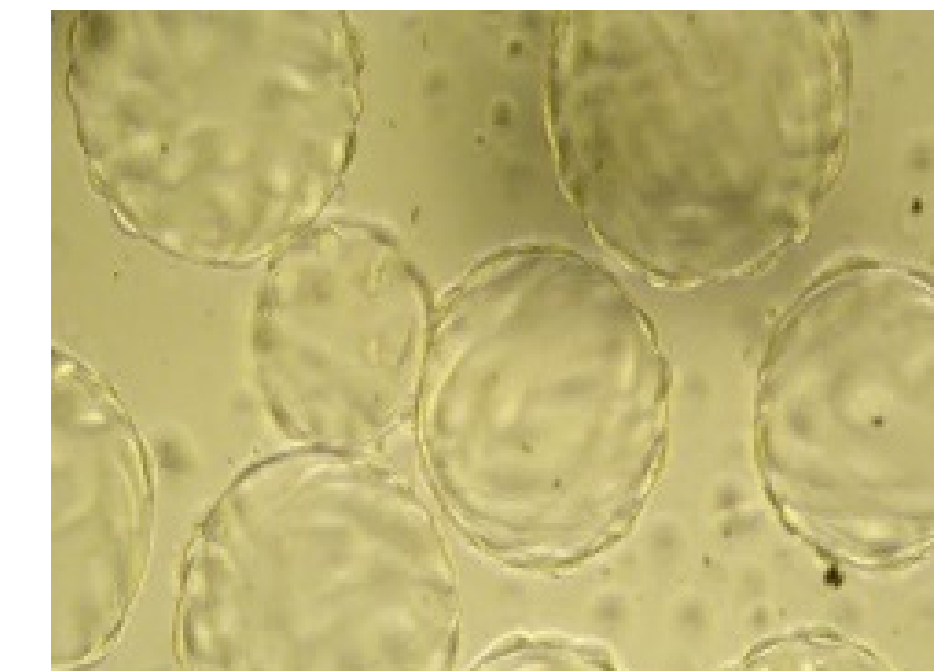
**Incubation:** 37°C with 5% CO<sub>2</sub> v/v

The spinner flask was fed when the concentration of glucose dropped below 0.5 g/L by allowing the microcarriers to settle down

## FRhK-4 cells on microcarriers

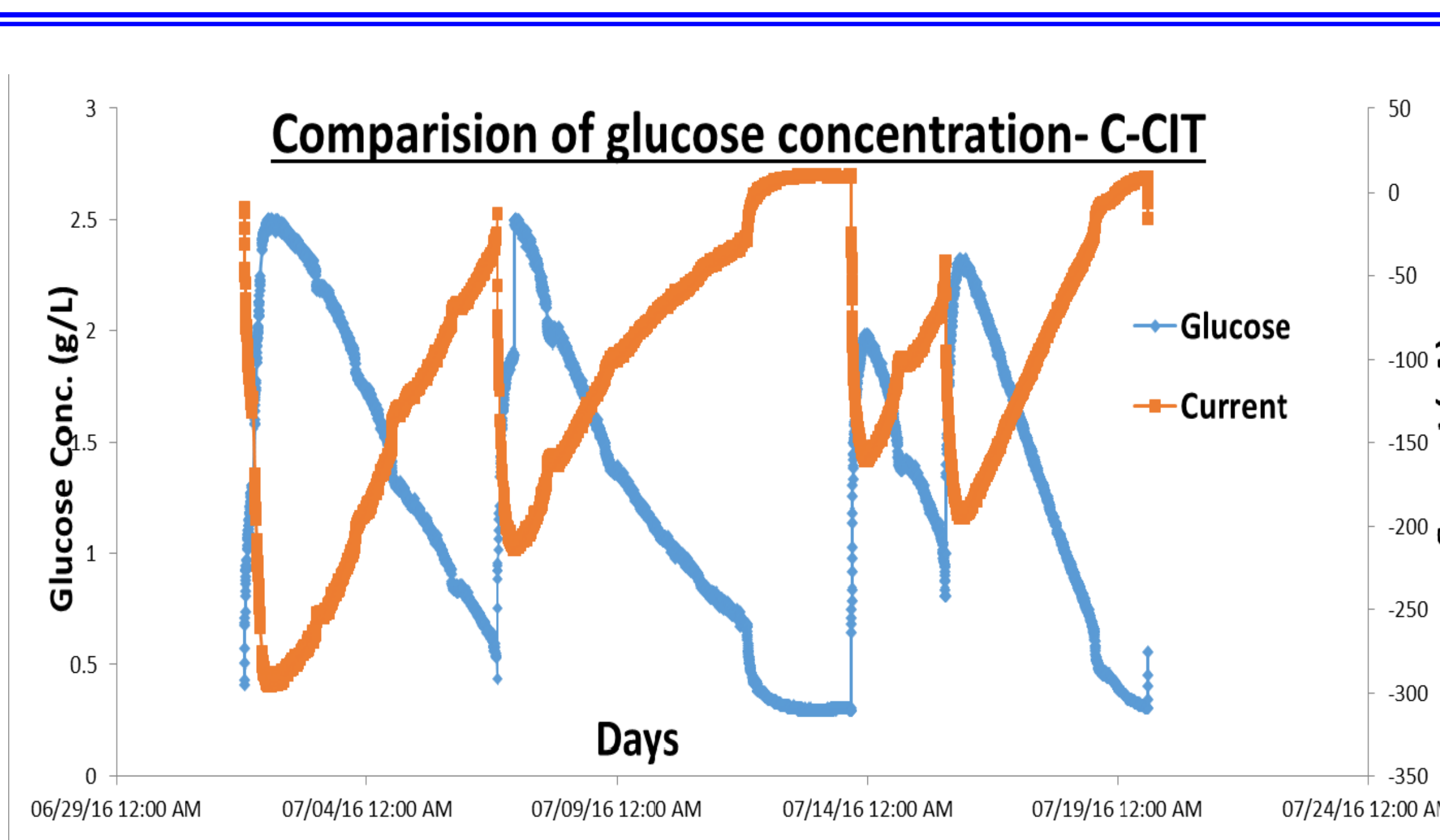


Day 0



Day 7

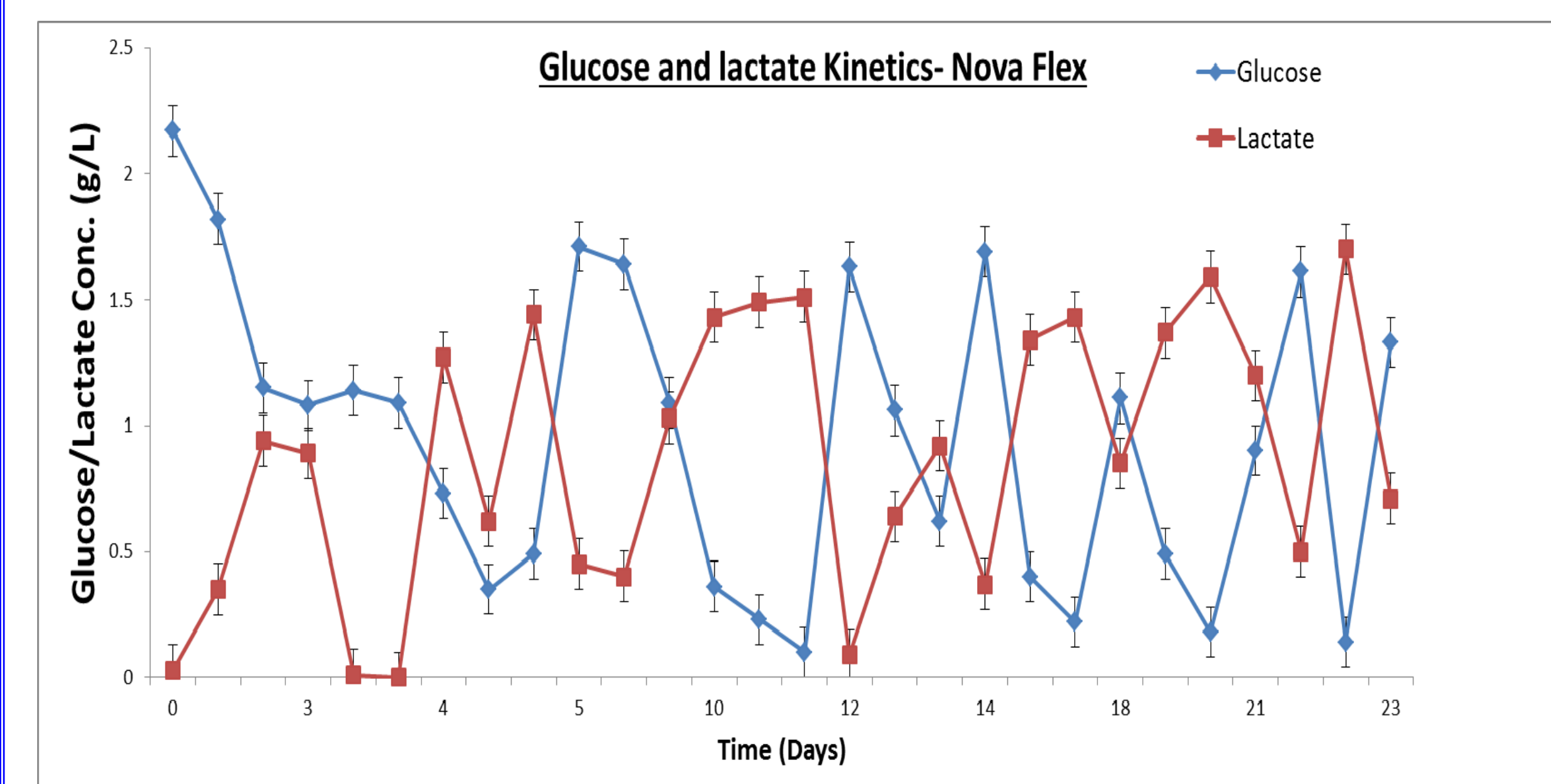
## RESULTS



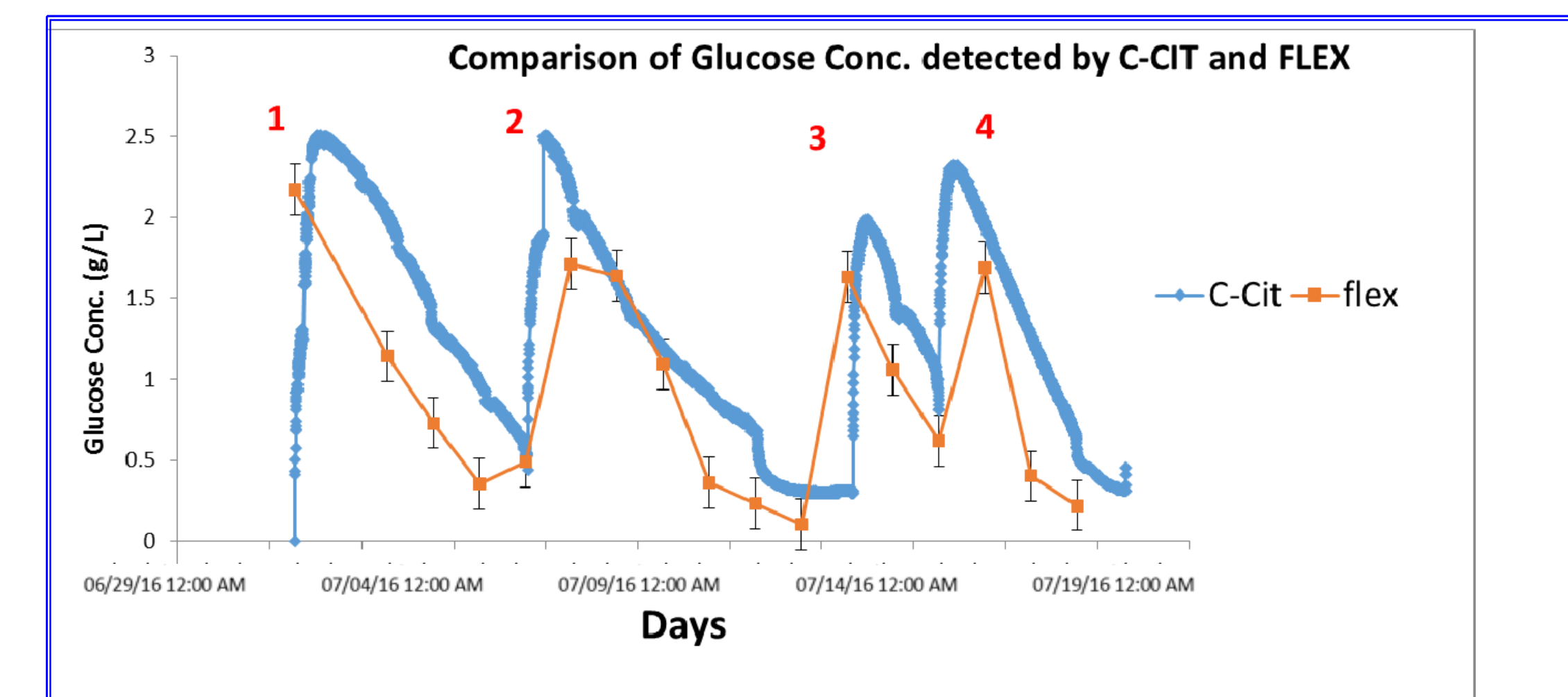
**Figure 1:** In-situ glucose measurement was performed for 22 days in the spinner flask with FRhK-4 cells. (data collected before and after feeding)

### Notes:

- Current data (nA) in the above Fig.1, is a measurement of electrons produced during the oxidation process (glucose with immobilized glucose oxidase on sensor)
- C-CIT software acts as an interface to convert the current (nA) to glucose concentration (g/L)



**Figure 2:** Comparison of glucose and lactate concentrations with offline analyzer Nova-FLEX (feeding was carried out at different time points over 22 days)



**Figure 3:** Comparison of glucose concentration with offline analyzer (FLEX) and online sensor (C-CIT) (data collected during 22 days culture process with four feeding points)

## CONCLUSION

- ❖ Decrease and increase in glucose levels were correctly detected before and after feeding, during the entire cell culture process
- ❖ Glucose trends with the online sensor and offline analyzer were similar
- ❖ *In situ* online sensor lasts long enough to support both batch and fed-batch processes

## FUTURE WORK

- ❖ Include HPLC as offline method for comparing glucose concentration data with that from the online sensor
- ❖ Optimize protocol: shorten conditioning and calibration processes (automatically by software) to facilitate combining medium, cells and sensor, in one step
- ❖ Study the effect of current (nA) vs. agitation temperature and magnetic field on glucose concentration
- ❖ Assess contamination risks to cell culture when using online sensor technology
- ❖ Integrate (C-CIT Sensors) technology into microbial culture processes

## ACKNOWLEDGEMENT

C-CIT Sensors AG, Switzerland and Nova Biomedical Corporation, USA, for support